

TransforMax™ EPI300™ Electrocompetent *E. coli*

TransforMax™ EPI300™ Chemically Competent *E. coli*

Cat. Nos. EC300105, EC300110, EC300150, and C300C105

TransforMax™ EPI300™ *E. coli* have been specifically engineered for use with Epicentre's CopyControl™ Cloning Systems.* The cells contain a mutant *trfA* gene, whose protein product is required for initiation of replication from the *oriV* origin of replication contained on the CopyControl vectors and on clones retrofitted with an EZ-Tn5™ <*oriV*/KAN-2> Transposon. The *trfA* gene is under tightly regulated control of an inducible promoter. When grown on standard LB plates or in LB or SOC culture medium, expression of the *trfA* gene is repressed. Addition of the CopyControl Induction Solution (provided with the cells) induces expression of the *trfA* gene and subsequent utilization of the *oriV* origin of replication and high copy amplification of the CopyControl BAC, fosmid and PCR clones.

Important Phenotypes and Applications

- Mutant *trfA* gene under tightly regulated control of an inducible promoter for complete copy number control of CopyControl clones.
- Supports blue/white screening of vectors expressing the LacZ' α -complementing peptide.
- Restriction minus for efficient cloning of methylated genomic (e.g. mammalian genomic) DNA.
- Endonuclease minus (*endA1*) to ensure high yields of clones.
- Recombination minus (*recA1*) to ensure the stability of large cloned inserts.

Cat. #	Quantity
TransforMax™ EPI300™ Electrocompetent <i>E. coli</i> are available in three sizes:	
EC300105	5 x 100 μ l
EC300110	10 x 100 μ l
EC300150 (shipped as 5 x EC300110)	50 x 100 μ l

TransforMax™ EPI300™ Chemically Competent *E. coli*:

C300C105 (10 Transformations)	10 x 50 μ l
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Each is supplied with CopyControl™ Induction Solution and pUC19 Control DNA (100 μ g/ μ l).

Related Products: The following products are also available:

- CopyControl™ BAC Cloning Kits
- CopyControl™ PCR Cloning Kit
- EZ-Tn5™ <oriV/KAN-2> Insertion Kit
- CopyControl™ Induction Solution

Product Specifications:

Storage: Store the TransforMax EPI300 *E. coli* cells at -70°C . The CopyControl Induction Solution and the pUC19 Control DNA can be stored at either -20°C or -70°C . Warm the CopyControl Induction Solution to room temperature and mix thoroughly before use.

Genotype:

F⁻ *mcrA* Δ (*mrr-hsdRMS-mcrBC*) ϕ 80d*lacZ* Δ M15 Δ *lacX74 recA1 endA1 araD139 Δ (*ara, leu*)7697 *galU galK* λ^{-} *rpsL nupG trfA dhfr*.*

Quality Control: TransforMax EPI300 Electrocompetent *E. coli* have a transformation efficiency of $>1 \times 10^{10}$ cfu/ μg DNA using 10 pg of pUC19 and an Eppendorf Multipipetator (2.5 kV, fast charge rate) using a 2 mm cuvette. TransforMax EPI300 Chemically Competent *E. coli* have a transformation efficiency of $>5 \times 10^8$ cfu per microgram of DNA using 10 pg of pUC19. Both electrocompetent and chemically competent cells are tested for induced expression of the *trfA* gene using transformants harboring a plasmid with an *oriV* origin of replication, and to be free of contaminating DNA rendering resistance to ampicillin, tetracycline, kanamycin and chloramphenicol.

Electroporation of TransforMax EPI300 Electrocompetent *E. coli*

Note: The electroporation procedure described here uses 50 μl of electrocompetent cells. A different volume of cells can also be used based on the experiences and needs of the user.

1. DNA should be in water or very low salt buffer (e.g. TE Buffer: 10 mM Tris-HCl [pH 7.5], 1 mM EDTA) to prevent arcing during electroporation. The pUC19 Control DNA is provided in TE at 100 pg/ μl . If running a transformation control, dilute the pUC19 Control DNA 1:10 (to a final concentration of 10 pg/ μl) with sterile, deionized water and use 1 μl for electroporation.
2. Prepare 1 ml of SOC or LB medium (**do not** include antibiotic in the medium) for each electroporation to be performed. This medium will be used for post-electroporation outgrowth of transformed cells. Maintain the medium at room temperature.
3. Pre-chill electroporation cuvettes and 1.5 ml tubes on ice.
4. Set-up the electroporation device according to the manufacturer's recommendations for bacterial (*E. coli*) electroporation.
5. Thaw TransforMax EPI300 Electrocompetent *E. coli* cells on ice. Mix gently. Use the cells immediately. Unused cells can be refrozen at -70°C .

Note: *Refrozen cells may have reduced transformation efficiency, but for most cases, the reduction is not significant enough to interfere with the desired results.*

6. Transfer the desired amount of DNA and 50 µl of cells to a pre-chilled microcentrifuge tube.

Note: *a smaller volume of cells can be used based on the needs and experiences of the user. Mix the cells and DNA by pipetting up and down 2-3 times.*

7. Transfer the cell/DNA mix to the electroporation cuvette. Be sure that there are no air bubbles in the cuvette. Wipe the cuvette of any condensation. Place into the electroporator and apply the electric pulse at the manufacturer's recommendations for bacterial (*E. coli*) electroporation.
8. Immediately after electroporation, add 950 µl of room temperature SOC medium [Hanahan, D., (1983) *J. Mol. Biol.*, **166**, 557] to the cuvette. Mix gently by pipetting up and down 2-3 times.
9. Transfer the cells to a 15 ml tube and incubate at 37°C with shaking at 220-230 rpm for 1 hour to recover the cells and allow expression of the antibiotic resistance marker.
10. Dilute and plate the cells on appropriate medium and antibiotic. For cells transformed with the control pUC19 DNA, plate on LB agar containing 100 µg/ml of ampicillin. The remaining cell outgrowth can be stored at 4°C in the event additional cell dilutions are plated.

Transformation of TransforMax EPI300 Chemically Competent *E. coli*

This procedure was written for the transformation of 50 µl of TransforMax EPI300 Chemically Competent *E. coli*. The procedure can be scaled down as needed.

1. Prepare 250 µl of SOC medium (**do not** include antibiotic in the medium) for each transformation to be performed. Maintain the medium at room temperature.
2. Chill 1.5 ml microcentrifuge tubes or microplate at 4°C and heat a water bath or other temperature-controlled apparatus to 42°C.
3. Thaw TransforMax EPI300 Chemically Competent *E. coli* cells on ice. Mix by *gentle* tapping. Use the cells immediately.

Note: *Refreezing chemically competent cells will result in a greatly reduced transformation efficiency.*

4. Transfer 1-5 µl of DNA and 50 µl of cells to a pre-chilled, microcentrifuge tube or into the wells of a microplate. Cover the cells and incubate on ice for 30 minutes.
5. Transfer the tubes or plate to 42°C and heat shock for 30 seconds.
6. Transfer the cells back to ice and cool for 2 minutes.
7. Add 250 µl of the SOC medium to each tube.
8. Recover the cells by incubating at 37°C for 60 minutes with shaking at 220-230 rpm.
9. Plate the cells on the appropriate media and antibiotic.

Induction of CopyControl Fosmid, CopyControl PCR Clones and Fosmids Retrofitted with an EZ-Tn5 <oriV /KAN-2> Transposon to High Copy Number

CopyControl Fosmid clones, CopyControl PCR clones and low copy number fosmid clones retrofitted with the EZ-Tn5 <oriV /KAN-2> Transposon and grown in TransforMax EPI300 cells can be amplified to 10-50 copies per cell. The induction process can be done in any culture volume desired depending on the needs of the user. Generally, a 1 ml induced culture will provide a sufficient amount of DNA for most applications including sequencing and fingerprinting. Here we provide the procedure for amplifying the clones in 1 ml, 5 ml and 50 ml cultures.

Important: *The Growth Media for amplifying CopyControl Fosmid clones and CopyControl PCR clones and low copy number fosmid clones retrofitted with an EZ-Tn5 <oriV/KAN-2> Transposon are different. Be sure to use the appropriate Growth Media for the clones that you are amplifying.*

Growth Media for CopyControl Fosmid Clones and CopyControl PCR Clones

LB + chloramphenicol (12.5 µg/ml)

Growth Media for Fosmid Clones Retrofitted with an EZ-Tn5 <oriV/KAN-2> Transposon

LB + chloramphenicol* (12.5 µg/ml) + kanamycin (50 µg/ml)

**or other selectable marker present on the cloning vector backbone*

1. Add 5 ml of the appropriate Growth Media to 15 ml tubes for each fosmid or PCR clone that will be induced to high copy number.
2. Individually inoculate the media with a small portion of the desired fosmid or PCR clones grown on an overnight plate.
3. Grow the cultures overnight at 37°C with shaking. These cultures will be used as inocula for the copy number amplification procedure.
4. From the table below, combine the appropriate volumes of fresh Growth Media, the overnight culture and the CopyControl Induction Solution for the desired volume of induction culture. Aeration of the induction cultures is critical. Therefore, to maximize the surface area of the culture solution in the tube, perform the induction in the largest volume tubes that reasonably meets your needs and resources. For example, induce clones to high copy number in 1 ml of culture, using 1.5 ml tubes or larger, 5 ml cultures in 15 ml tubes and 50 ml cultures in 125 ml flasks.

Total volume of clone induction culture	Volume of fresh LB + chloramphenicol (12.5 µg/ml)	Volume of overnight 5 ml culture	Volume of 1000X CopyControl Induction Solution*
1 ml	800 µl	200 µl	1 µl
5 ml	4.5 ml	500 µl	5 µl
50 ml	45 ml	5 ml	50 µl

*Mix thoroughly after thawing.

- Vigorously shake the tubes at 37°C for 5 hours. Aeration is critical! Shake the tubes in a manner that will maximize aeration of the cultures (for example 1.5 ml tubes can be taped horizontally to the shaking table).
- Centrifuge the cells and purify the DNA by your standard lab methods.

Induction of CopyControl BAC Clones and BAC Clones Retrofitted with an EZ-Tn5 <oriV/KAN-2> Transposon to High Copy Number

CopyControl BAC clones and low copy number BAC clones retrofitted with an EZ-Tn5 <oriV/KAN-2> Transposon and grown in TransforMax EPI300 cells can be amplified to 10-20 copies per cell. Generally, 1 ml of an induced culture will provide a sufficient amount of BAC DNA for most applications including sequencing and fingerprinting. Procedures for amplifying CopyControl BAC clones and low copy number BAC clones retrofitted with an EZ-Tn5 <oriV/KAN-2> Transposon in 1.5 ml tubes and in deep-well, 96 well plates are provided. The induction volumes can be scaled up as required by the user.

Important: The Growth Media for amplifying CopyControl BAC clones and low copy number BAC clones retrofitted with an EZ-Tn5 <oriV/KAN-2> Transposon are different. Be sure to use the appropriate Growth Media for the type of BAC clones that you are amplifying.

Growth Media for CopyControl BAC Clones

LB + chloramphenicol (12.5 µg/ml)

Growth Media for BAC Clones Retrofitted with an EZ-Tn5 <oriV/KAN-2> Transposon

LB + chloramphenicol* (12.5 µg/ml) + kanamycin (50 µg/ml)

*or other selectable marker present on the cloning vector backbone

Amplification of BAC clones in 1.5 ml tubes

1. Dispense 1 ml of the appropriate Growth Media into 1.5 ml tubes. Inoculate each tube with an isolated single BAC clone from an overnight plate.
2. Incubate the cultures at 37°C overnight without shaking.
3. Following overnight incubation, mix each tube and then aspirate off 800 µl of culture medium from each and discard.
4. Add 800 µl of fresh Growth Media into each tube containing the remaining 200 µl of the overnight culture. Mix by vortexing.
5. Incubate the tubes for 30 minutes at 37°C with shaking at 250 rpm. After 30 minutes, the O.D₆₀₀ will be at 0.4 - 0.6.
6. Thaw the CopyControl Induction Solution and mix thoroughly. Add 1 µl of 1000X CopyControl Induction Solution (to a 1X final concentration) to each tube. Incubate each for 2 hour at 37°C with vigorous shaking. Aeration is critical! Shake the tubes in a manner that will maximize aeration of the cultures (for example 1.5 ml tubes can be taped horizontally to the shaking table).
7. Isolate DNA from the induced culture by your method of choice.

Amplification of CopyControl BAC clones in deep-well (2 ml) 96 well plates

1. Dispense 1 ml of the appropriate Growth Media into each well of a deep-well plate. Inoculate each well with an isolated single BAC clone from an overnight plate.
2. Seal the plate with porous microtiter plate sealer, incubate at 37°C overnight without shaking.
3. Following overnight incubation, mix the cultures by shaking and then aspirate off 800 µl of culture medium from each well and discard.
4. Add 800 µl of fresh Growth Media into each well containing the remaining 200 µl of the overnight culture. Mix by shaking or vortexing.
5. Incubate the plate for 30 minutes at 37°C with shaking at 250 rpm. After 30 minutes, the O.D₆₀₀ will be at 0.4 - 0.6.
6. Thaw the CopyControl Induction Solution and mix thoroughly. Add 1 µl of 1000X CopyControl Induction Solution (to a 1X final concentration) to each well. Incubate each for 2 hour at 37°C with vigorous shaking. Aeration is critical!
7. Isolate DNA from the induced culture by your method of choice.

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726 Post Road, Madison, WI 53713 (800) 284-8474 (608) 258-3080 Fax (608) 258-3088