

# TargetAmp™ 1-Round Biotin-aRNA Amplification Kit 105

Cat. No. TAB1R80510 – 10 Reactions

Cat. No. TAB1R80524 – 24 Reactions

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## 1. Introduction

### Features and Benefits of the TargetAmp 1-Round Biotin-aRNA Amplification Kit 105:

- 1) Produces microgram amounts of Biotin-aRNA (Biotin-cRNA) from as little as 25 ng of total RNA.
- 2) Linear RNA amplification process that preserves the relative transcript abundance of the sample.
- 3) Virtually eliminates template-independent reactions.
- 4) Reproducible amplification results.
- 5) Single tube reaction, no need to purify the cDNA transcription template prior to the *in vitro* transcription reaction.
- 6) Fast reaction times...completed in about 6 hours.
- 7) Easy to use...color-coded tubes and a simplified pipetting scheme reduce labor and the possibility of error.

The TargetAmp 1-Round Biotin-aRNA Amplification Kit 105 utilizes an improved linear RNA amplification procedure<sup>1</sup> to produce microgram amounts of biotin-labeled anti-sense RNA (biotin-aRNA; also called biotin-cRNA) from as little as 25 ng of total cellular RNA. The kit provides optimal results when using SuperScript III Reverse Transcriptase (Invitrogen; provided by the user) in the reverse transcription step. The complete TargetAmp 1-Round Biotin-aRNA Amplification Kit 105 procedure can be completed in about 6 hours (Fig. 1).

- A. First-strand cDNA Synthesis:** The Poly(A) RNA component of a total RNA sample is reverse transcribed into first strand cDNA. The reaction is primed from a synthetic oligo(dT) primer containing a phage T7 RNA Polymerase promoter sequence at its 5' end. First strand cDNA synthesis is catalyzed by SuperScript III Reverse Transcriptase (provided by the user) and performed at an elevated temperature to reduce RNA secondary structure.
- B. Second-strand cDNA Synthesis:** The cDNA produced is converted to double-stranded DNA (dsDNA) containing a T7 transcription promoter in an orientation that that will generate anti-sense RNA (aRNA; also called cRNA) during the subsequent *in vitro* transcription reaction. The dsDNA produced can be used in the *in vitro* transcription reaction without the need for purification.
- C. In Vitro Transcription of Biotin-aRNA:** High yields of Biotin-aRNA (Biotin-cRNA) are produced in a rapid *in vitro* transcription reaction that utilizes the double-stranded cDNA produced as template.

### Performance Specifications and Quality Control

The TargetAmp 1-Round Biotin-aRNA Amplification Kit 105 is function-tested in a control reaction. The kit must produce at least 8 µg of Biotin-aRNA (cRNA) from 80 ng of HeLa Total RNA Control, corresponding to a greater than 5,000-fold amplification of the Poly(A) RNA assuming that 2% of the HeLa Total RNA Control is Poly(A) RNA. A negative control reaction ("no-RNA" control) produces less than 1 µg of Biotin-aRNA.

## 2. Kit Contents

The kit components are supplied in tubes with colored caps for easier identification.

The kit has been developed for use with and will provide optimal results with SuperScript® III Reverse Transcriptase (Invitrogen Corp.; provided by the user).

Component Name	Tube Label	10 Reactions	24 Reactions	Cap Color
TargetAmp T7-Oligo(dT) Primer A	T7-Oligo(dT) Primer A	10 µl	30 µl	Red
TargetAmp Reverse Transcription PreMix	RT PreMix	15 µl	50 µl	
TargetAmp DNA Polymerase PreMix	DNA Pol. PreMix	50 µl	125 µl	
TargetAmp DNA Polymerase	DNA Polymerase	10 µl	18 µl	
TargetAmp T7 RNA Polymerase	T7 RNA Polymerase	20 µl	60 µl	Green
TargetAmp T7 Transcription Buffer	T7 Transcription Buffer	20 µl	60 µl	
TargetAmp NTP PreMix	NTP PreMix	100 µl	270 µl	
TargetAmp UTP / Biotin-UTP	UTP / Biotin-UTP	30 µl	90 µl	
RNase-Free DNase I	RNase-Free DNase	20 µl	55 µl	
100 mM Dithiothreitol (DTT)	DTT	50 µl	100 µl	Clear
HeLa Total RNA Control (40 ng/µl)	HeLa Total RNA Control	10 µl	10 µl	
RNase-Free Water	RNase-Free Water	500 µl	500 µl	

**Storage:** Upon receipt of this kit, remove the tube containing the HeLa Total RNA Control and store it at  $-70^{\circ}\text{C}$  to  $-80^{\circ}\text{C}$ . Store the remainder of the kit at  $-20^{\circ}\text{C}$ .

### Additional Required Reagents and Equipment:

SuperScript III Reverse Transcriptase (Invitrogen Corp.)

Thermocycler or water bath

Microcentrifuge

RNase-Free Water

RNeasy® MinElute® Cleanup Kit (Qiagen) or RNeasy Mini Kit (Qiagen) (see "Biotin-aRNA Purification")

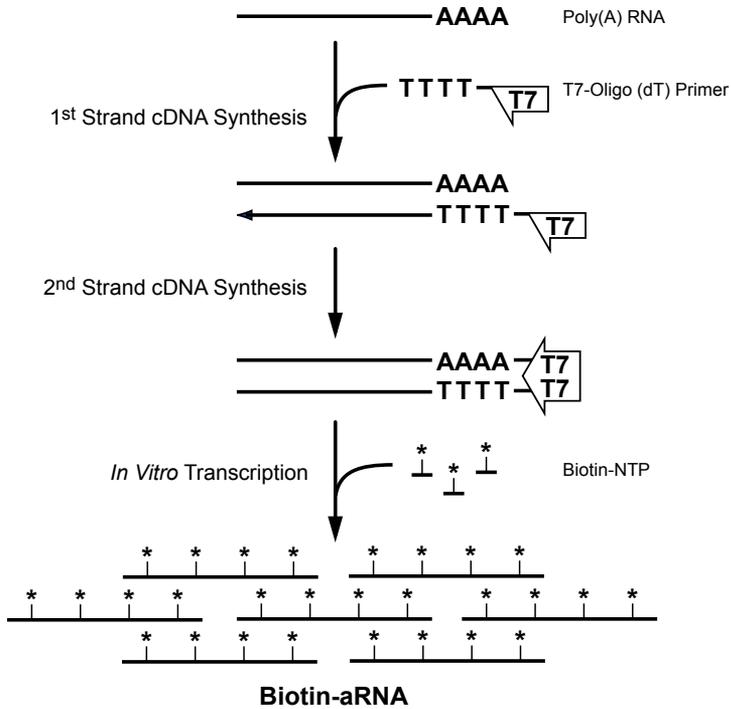


Figure 1. TargetAmp™ 1-Round Biotin-aRNA Amplification Kit 105 Procedure.

### 3. Related Products

#### Kits for Isolating Total Cellular RNA

##### ArrayPure™ Nano-scale RNA Purification Kit

MPS04050

50 Purifications

The ArrayPure Kit provides all reagents needed to purify total RNA from 1-10,000 eukaryotic cells without the use of organic solvents such as phenol or columns.

##### MasterPure™ RNA Purification Kit

MCR85102

100 Purifications

The MasterPure RNA Purification Kit provides all reagents needed to purify total RNA from >10,000 eukaryotic cells without the use of organic solvents such as phenol or columns.

## 4. Preparation

### Assessing the Quality of the Total RNA

The success of microarray experiments is strongly influenced by the quality of the RNA. RNA quality has two components: purity of the RNA (or absence of contaminants) and integrity (intactness) of the RNA. RNA quality should be assessed prior to every RNA amplification reaction. Poor quality RNA is the most common cause of sub-optimal RNA amplification results!

RNA Purification Methods and RNA Purity. Total cellular RNA, isolated by a number of methods, can be amplified successfully using the TargetAmp 1-Round Biotin-aRNA Amplification Kit 105. However, it is very important that the purified RNA be free of salts, metal ions, ethanol, and phenol which can inhibit the enzymatic reactions performed in the RNA amplification process. Commonly used RNA extraction and purification methods that are compatible with the TargetAmp RNA amplification process include but are not limited to:

**TRIzol®/TRI Reagent®**, a homogeneous solution of the powerful denaturants guanidinium isothiocyanate and phenol, is very effective at extracting the RNA from the cells. However all traces of guanidinium salts and phenol must be removed from the RNA sample prior to the RNA amplification process. If you precipitate the RNA from TRIzol-extracted cells, be sure to wash the RNA pellet at least two times with cold 70-75% ethanol to remove all traces of phenol and guanidinium salts. Air dry the RNA pellet (do not use a vacuum centrifuge) to remove residual ethanol. Then, resuspend the RNA in RNase-Free water. If you purify the RNA from TRIzol-extracted cells by column purification methods, please read the section "Spin Columns" immediately following.

**Spin Columns** (e.g., the RNeasy MinElute Cleanup Kit and RNeasy Mini Kit from Qiagen) are effective in purifying RNA samples that are free of the contaminants that may inhibit the RNA amplification process. Spin columns can be used with most RNA extraction procedures (e.g. TRIzol reagent). If using spin columns, follow the manufacturer's instructions closely. Make sure to remove all washing solution prior to the RNA elution step. Then, elute the RNA from the column membrane using RNase-Free water.

**Salt-Fractionation:** RNA purification that employs gentle salt-fractionation, such as Epicentre's MasterPure™ RNA Purification Kit and ArrayPure™ Nano-scale RNA Purification Kit, routinely produce the highest yield of intact RNA without the use of phenol, guanidinium salts or columns. The ArrayPure kit has been developed for total RNA purification from 1-10,000 cells obtained by laser-capture methods such as Laser Capture Microdissection (LCM), from biopsy samples, from cell culture, or quick-frozen tissue. To purify RNA from >10,000 cells, Epicentre's MasterPure RNA Purification Kit is recommended. When using these kits, be sure to wash the RNA pellet at least two times with cold 70-75% ethanol to remove all traces of salts. Air dry the RNA pellet (do not use a vacuum centrifuge) to remove residual ethanol. Then, resuspend the RNA in RNase-Free water. The ArrayPure kit and MasterPure kit are not recommended for purification of RNA from tissue samples preserved with RNA<sub>later</sub>® or RNA<sub>later</sub>-ICE.

**RNA Integrity.** Successful microarray analysis using amplified RNA is dependent on an RNA sample that contains full-length, intact Poly(A) RNA. The most commonly used methods for assaying RNA integrity are by denaturing agarose gel electrophoresis or using an Agilent 2100 Bioanalyzer.

The advantages of denaturing agarose gel electrophoresis are its low cost and ready availability of the reagents required. Denaturing gel electrophoresis separates the RNAs by size (electrophoretic mobility) under denaturing conditions. Denaturing conditions are necessary to eliminate inter- and intra-molecular interactions within the RNA sample which may cause abnormal electrophoretic mobility. Following electrophoresis, the denaturing gel is stained with, for example, ethidium bromide and the user looks for the highly stained 18S and 28S rRNAs. These bands should be sharp and discrete with an absence of smearing under either. Based on these visual observations, the user infers that the mRNA in the sample is equally intact (or degraded). If the rRNA bands appear degraded, as evidenced by smearing under each, the RNA sample should be discarded and a new sample of total RNA purified. Ideally, the ethidium bromide stained 28S rRNA band should appear to be about twice as intense as the 18S rRNA band. The main disadvantage of denaturing gels is that they require up to 1 µg of total RNA be loaded per lane...a quantity of RNA that often is not expendable for those performing RNA amplification.

The Agilent 2100 Bioanalyzer is currently the preferred method for evaluating the integrity of an RNA sample. Like a denaturing gel, the bioanalyzer separates the RNAs by size (electrophoretic mobility). However, in contrast to a denaturing gel, the 2100 Bioanalyzer requires nanogram amounts of total RNA per well when using the manufacturer's RNA 6000 Nano LabChip®. When analyzing the RNA sample using the Agilent 2100 Bioanalyzer, the 18S and 28S rRNA species should appear as distinct, sharp peaks on the electropherogram. A slightly increased baseline, indicative of the 1-5% Poly(A) RNA contained in the sample, can be seen between the two peaks.

### **Maintaining an RNase-free Environment**

Ribonuclease contamination is a significant concern for those performing RNA amplification. The ubiquitous RNase A is a highly stable and active ribonuclease that can contaminate any lab environment and is present on human skin. All components of the TargetAmp 1-Round Biotin-aRNA Amplification Kit 105 have been tested to ensure the lack of contaminating ribonuclease activities. However, creating an RNase-free work environment and maintaining RNase-free solutions is critical for performing successful RNA amplification reactions. Therefore, we strongly recommend that the user:

- 1) Use RNase-free or autoclaved tubes and pipette tips
- 2) Always wear gloves when handling samples containing RNA. Change gloves frequently especially after touching potential sources of RNase contamination such as door knobs, pens, pencils and human skin.
- 3) Always wear gloves when handling kit components. Do not pick up any kit component with an ungloved hand.
- 4) Keep all kit components tightly sealed when not in use. Keep all tubes containing RNA tightly sealed during the incubation steps.

## Input Total RNA and Biotin-aRNA Yield

The TargetAmp 1-Round Biotin-aRNA Amplification Kit 105 is extremely efficient at amplifying very small quantities of Poly(A) RNA in a total RNA Preparation. The TargetAmp kit has been evaluated for production of Biotin-aRNA from as little as 25 ng of total RNA using the HeLa Total RNA control provided in the kit. User results may vary depending on the quality of the total RNA sample and the Poly(A) content of the sample.

**Note:** To amplify <25 ng of total RNA use the TargetAmp 2-Round Biotin-aRNA Amplification Kit 3.0.

**Table 1. Yields of Biotin-aRNA (Biotin-cRNA) obtained using the TargetAmp™ 1-Round Biotin-aRNA Amplification Kit 105.** Results summarize experiments performed using the HeLa Total RNA control provided in the kit.

HeLa Control total RNA	HeLa Poly(A) RNA [assume 2% of HeLa total RNA is Poly(A) RNA]	Biotin-aRNA Yield	Fold-Amplification of HeLa Poly(A) RNA [Biotin-aRNA yield / Poly(A) content]
25 ng	0.5 ng	3.1 µg (3100 ng)	6,200
50 ng	1 ng	5.3 µg (5,300 ng)	5,300
100 ng	2 ng	10.8 µg (10,800 ng)	5,600
200 ng	4 ng	22.6 µg (22,600 ng)	5,650
400 ng	8 ng	47.2 µg (47,200 ng)	5,900
500 ng	10 ng	75.6 µg (75,600 ng)	7,500

**Important!** The TargetAmp 1-Round Biotin-aRNA Amplification Kit 105 has been optimized for producing >5,000-fold amplification of the Poly(A) RNA from 25-500 ng of total cellular RNA per reaction. Amplifying >500 ng of total RNA in a single TargetAmp 1-Round Biotin-aRNA Amplification Kit 105 reaction may result in less than 5,000-fold amplification of the Poly(A) RNA and may result in under-representation of some Poly(A) RNA sequences in the Biotin-aRNA produced. Therefore, if it is desirable to perform an amplification reaction using >500 ng of total RNA, we strongly recommend that the user perform multiple reactions, each containing up to but not exceeding 500 ng of total RNA.

## Additional Suggestions

### Familiarize Yourself with the TargetAmp 1-Round Biotin-aRNA Amplification Kit 105 Procedure

The TargetAmp 1-Round Biotin-aRNA Amplification Kit 105 includes many reagents. Before starting, please read this protocol carefully and familiarize yourself with each kit component and in which step of the RNA amplification process it is used. Be sure to wear gloves when handling the kit components.

## Importance of Running the Control Reaction

We strongly recommend that those who are not experienced with the TargetAmp 1-Round Biotin-aRNA Amplification Kit 105 perform a control amplification reaction (Appendix 1) prior to committing a precious sample. HeLa total RNA is provided with the kit as a control.

## Performing the TargetAmp 1-Round Biotin-aRNA Amplification Kit 105 Reactions

We recommend that all reactions be performed in sterile 0.2 ml or 0.5 ml thin-walled tubes using sterile pipette tips and recently calibrated pipettors. Very small volumes of some kit components are required for each reaction. Therefore, we recommend the user prepare Master Mixes of reaction components when amplifying multiple samples.

### Some Simple but Important Factors for Obtaining Optimal Results

- 1) Familiarize yourself with the kit by running a control reaction (Appendix 1) before committing a precious sample.
- 2) Use up to but not more than 500 ng of total RNA per reaction.
- 3) Assemble the *in vitro* transcription reaction (Step C) at room temperature.  
Do not exceed the *in vitro* transcription reaction time indicated in the procedure.
- 4) Optional stopping points are noted following the 2<sup>nd</sup>-strand cDNA synthesis step (Part B) and after purification of the Biotin-aRNA (Part D).

## 5. RNA Amplification Procedure

Please read through the TargetAmp 1-Round Biotin-aRNA Amplification Kit 105 procedure carefully before beginning. We strongly recommend that those who are not experienced with the TargetAmp kit perform a control amplification reaction (Appendix 1) prior to committing a precious sample. A HeLa total RNA control is provided with the kit.

An abbreviated (single page) protocol is presented on page 15 for users experienced in using this kit.

### A. First-strand cDNA Synthesis

SuperScript III Reverse Transcriptase (Invitrogen) is required for use in this Step. The SuperScript III enzyme is provided by the user. The total RNA sample must be free of contaminating salts, metal ions, ethanol, and phenol. For best results, the RNA sample should be dissolved in RNase-Free water.

Required in Part A

Component Name	Tube Label	Cap Color
TargetAmp T7-Oligo(dT) Primer A	T7-Oligo(dT) Primer A	Red
TargetAmp Reverse Transcription PreMix	RT PreMix	
Dithiothreitol	DTT	Clear
RNase-Free Water	RNase-Free Water	

**Incubation temperatures performed in Step A:** 50°C and 65°C.

**Important!** The TargetAmp 1-Round Biotin-aRNA Amplification Kit 105 has been optimized for producing >5,000-fold amplification of the Poly(A) RNA from 25-500 ng of total cellular RNA per reaction. Amplifying >500 ng of total RNA in a single TargetAmp 1-Round Biotin-aRNA Amplification Kit 105 reaction may result in less than 5,000-fold amplification of the Poly(A) RNA and may result in under-representation of some Poly(A) RNA sequences in the Biotin-aRNA produced. Therefore, if it is desirable to perform an amplification reaction using >500 ng of total RNA, we strongly recommend that the user perform multiple reactions, each containing up to but not exceeding 500 ng of total RNA.

- Anneal the TargetAmp T7-Oligo(dT) Primer A to the RNA sample. If a “no template” control reaction is performed, substitute 2 µl of RNase-Free Water for the Total RNA sample.

x µl	RNase-Free Water
x µl	Total RNA sample (25-500 ng)
1 µl	TargetAmp T7-Oligo(dT) Primer A
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3 µl	Total reaction volume

- Incubate at 65°C for 5 minutes in a water bath or thermocycler.
- Chill on ice for 1 minute. Centrifuge briefly in a microcentrifuge.
- Prepare the 1<sup>st</sup>-Strand cDNA Synthesis Master Mix.

For each 1<sup>st</sup>-strand cDNA synthesis reaction, combine on ice:

1.5 µl	TargetAmp Reverse Transcription PreMix
0.25 µl	DTT
0.25 µl	SuperScript III Reverse Transcriptase (200 U/µl)
<hr/>	
2 µl	Total reaction volume

**Important!** Be sure to use SuperScript III Reverse Transcriptase. Do not use the SuperScript 5X Buffer or the DTT that is provided with the enzyme.

- Gently mix the 1<sup>st</sup>-Strand cDNA Synthesis Master Mix and then add 2 µl of it to each reaction.
- Gently mix the reactions and then incubate each at 50°C for 30 minutes in a water bath or thermocycler. If the thermocycler has a heated lid function, heat the lid only if the temperature of the lid can be maintained at about 50°C.

**B. Second-strand cDNA Synthesis**

Required in Part B

Component Name	Tube Label	Cap Color
TargetAmp DNA Polymerase PreMix	DNA Pol. PreMix	Red
TargetAmp DNA Polymerase	DNA Polymerase	

**Incubation temperatures performed in Step B:** 65°C and 80°C.

- Prepare the 2<sup>nd</sup>-Strand cDNA Synthesis Master Mix.  
For each 2<sup>nd</sup>-strand cDNA synthesis reaction, combine on ice:
 

4.5 µl	TargetAmp DNA Polymerase PreMix
0.5 µl	TargetAmp DNA Polymerase
5 µl	Total reaction volume
- Gently mix the 2<sup>nd</sup>-Strand cDNA Synthesis Master Mix and then add 5 µl of it to each reaction.
- Gently mix the reactions and then incubate at 65°C for 10 minutes in a water bath or thermocycler. Centrifuge briefly in a microcentrifuge.  
**Important!** Be sure to incubate the reactions at 65°C.
- Incubate the reactions at 80°C for 3 minutes.  
Centrifuge briefly in a microcentrifuge then chill on ice.  
**Note:** If desired, the reactions can now be frozen and stored overnight at -20°C.

### C. In Vitro Transcription of Biotin-aRNA

The *in vitro* transcription reaction produces Biotin-aRNA by incorporating Biotin-UTP into the RNA transcripts.

Required in Part C

Component Name	Tube Label	Cap Color
TargetAmp T7 RNA Polymerase	T7 RNA Polymerase	Green
TargetAmp T7 Transcription Buffer	T7 Transcription Buffer	
TargetAmp NTP PreMix	NTP PreMix	
TargetAmp UTP / Biotin-UTP	UTP / Biotin-UTP	
RNase-Free DNase I	DNase I	
Dithiothreitol	DTT	Clear
RNase-Free Water	RNase-Free Water	

#### Incubation temperatures performed in Step C: 37°C and 42°C.

- Warm the TargetAmp T7 RNA Polymerase to room temperature. Thaw the remaining *in vitro* transcription reagents at room temperature. If a precipitate is visible in the thawed TargetAmp T7 Transcription Buffer, heat the Buffer to 37°C until it dissolves. Keep the TargetAmp T7 Transcription Buffer at room temperature.
- Thoroughly mix the thawed T7 Transcription Buffer and the NTP PreMix solutions.  
**Important!** If a precipitate is visible in the thawed TargetAmp T7 Transcription Buffer, heat the Buffer to 37°C until it dissolves. Mix the Buffer thoroughly. Keep the Buffer at room temperature.

3. Prepare the *In Vitro* Transcription Master Mix.

For each *in vitro* transcription reaction, combine, in order, at room temperature:

2 µl	T7 Transcription Buffer
3 µl	UTP / Biotin-UTP
10 µl	NTP PreMix
3 µl	DTT
2 µl	T7 RNA Polymerase
<hr/>	
20 µl	Total reaction volume

4. Gently mix the *In Vitro* Transcription Master Mix and then add 20 µl of it to each reaction.

## 5. Gently mix the reactions and then incubate at 42°C for 4 hours in a thermocycler. If the thermocycler has a heated lid function, heat the lid only if the temperature of the lid can be maintained at about 42°C. If the lid temperature can not be maintained at about 42°C, then perform the incubations without heating the lid.

**Important!** Do not exceed 4 hour incubation at 42°C. Optimal yield and quality (length) of Biotin-aRNA is achieved in 4 hours.

## 6. Add 2 µl of RNase-Free DNase I to each reaction.

Mix gently and then incubate each at 37°C for 15 minutes.

**D. Biotin-aRNA Purification**

The purification column to use is dependent upon the expected yield of aRNA. Use the table on page 8 to estimate the yield of aRNA expected from the amount of total RNA used in each amplification reaction. Then,

*If the expected yield of aRNA is <40 µg:* purify the aRNA using the Qiagen RNeasy MinElute Cleanup Kit. (Qiagen cat. no. 74204)

*If the expected yield of aRNA is >40 µg:* purify the aRNA using the Qiagen RNeasy Mini Kit. (Qiagen cat. no. 74014)

Use the RNase-Free Water that is provided in the MinElute Cleanup Kit or the RNeasy Mini Kit. The following procedure should be used with either the MinElute Cleanup Kit or the RNeasy Mini Kit.

1. Prepare 350 µl of RLT/β-ME Solution for each sample. Combine the RLT/β-ME in the ratio of 1 ml of Buffer RLT (provided in the RNeasy kit) with 10 µl of β-ME (β-mercaptoethanol) as described in the RNeasy kit's handbook.
2. Prepare 650 µl of RPE Solution for each sample by diluting 1 volume of Buffer RPE (provided in the purification kits) with 4 volumes of 96-100% ethanol as described in the purification kit's handbook.
3. To each sample add:

48 µl	RNase-Free Water
350 µl	RLT/β-ME Solution
<hr/>	
250 µl	100% Ethanol

4. Apply each sample to the purification kit's spin column in a 2 ml collection tube. Centrifuge at  $>8,000 \times g$  for 15 seconds. Discard the flow-through.
5. Apply 650  $\mu\text{l}$  RPE Solution onto the column. Centrifuge at  $>8,000 \times g$  for 15 seconds. Discard the flow-through.
6. Apply 650  $\mu\text{l}$  80% ethanol onto the column. Centrifuge at  $>8,000 \times g$  for 15 seconds. Discard the flow-through.
7. Transfer the spin column into a new collection tube. Centrifuge at full speed for 5 minutes.
8. Transfer the spin column to a 1.5 ml collection tube. Elute the aRNA:  
If using the MinElute Cleanup Column, apply 20  $\mu\text{l}$  of RNase-Free Water directly onto the center of the silica-gel membrane. Wait for 1 minute. Centrifuge at full speed for 1 minute.  
If using the RNeasy Mini Column, apply 40  $\mu\text{l}$  of RNase-Free Water directly onto the center of the silica-gel membrane. Wait for 1 minute. Centrifuge at full speed for 1 minute.
9. If using the MinElute Cleanup Column, apply an additional 20  $\mu\text{l}$  of RNase-Free Water directly onto the center of the silica-gel membrane. Wait for 1 minute. Centrifuge at full speed for 1 minute.  
If using the RNeasy Mini Column, apply an additional 40  $\mu\text{l}$  of RNase-Free Water directly onto the center of the silica-gel membrane. Wait for 1 minute. Centrifuge at full speed for 1 minute.  
This second elution step recovers 25-35% of the final yield of biotin-aRNA.

**Note:** *If desired, reactions can now be quick frozen (e.g., dry ice/ethanol bath) and stored at  $-80^{\circ}\text{C}$ .*

## E. Quantifying the Concentration, Yield and Fold-Amplification of the Biotin-aRNA

**Concentration and yield:** The concentration of the Biotin-aRNA can be readily determined using a NanoDrop® ND-1000 UV-Vis Spectrophotometer available from NanoDrop Technologies. Alternately, due to the high yield of Biotin-aRNA that is produced by a TargetAmp reaction, the yield and concentration of Biotin-aRNA can be determined by standard UV spectroscopy.

1. Prepare a dilution of the Biotin-aRNA into the minimum volume of water or TE Buffer (10 mM Tris-HCl [pH 7.5], 1 mM EDTA) required by the spectrophotometer cuvette that will be used.
2. Zero the spectrophotometer at 260 nm using the diluents (water or TE buffer) that was used to dilute the Biotin-aRNA sample.
3. Measure and record the absorbance of the diluted Biotin-aRNA at 260 nm ( $A_{260}$ ).
4. Calculate the concentration of the Biotin-aRNA. Use the conversion factor that an  $A_{260}$  reading of 1.0 is equal to an RNA concentration of 40  $\mu\text{g}/\text{ml}$ .  
Biotin-aRNA concentration = ( $A_{260}$  reading)  $\times$  (dilution factor)  $\times$  (40  $\mu\text{g}/\text{ml}$ ).  
Example: Dilution for  $A_{260}$  measurement = 1:100 with an  $A_{260}$  of the 1:100 dilution = 0.15.  
Biotin-aRNA concentration = (0.15)  $\times$  (100)  $\times$  (40  $\mu\text{g}/\text{ml}$ ) = 600  $\mu\text{g}/\text{ml}$  = (0.6  $\mu\text{g}/\mu\text{l}$ ) Biotin-aRNA.

5. Calculate the yield of Biotin-aRNA using the formula:

Yield of Biotin-aRNA = (Biotin-aRNA Concentration) x (Volume of Biotin-aRNA).

Example: 50 µl of Biotin-aRNA recovered from column, 0.6 µg/µl Biotin-aRNA from in Part E. Step 4.

Biotin-aRNA yield = (0.6 µg/µl) x (50 µl) = 30 µg of Biotin-aRNA.

In this example, 1 µl of 0.6 µg/µl of Biotin-aRNA was used for the spectrophotometer reading so there are now 29.4 µg of Biotin-aRNA remaining.

**Fold amplification:** The fold-amplification of the reaction can be calculated once the yield of the Biotin-aRNA has been determined. However, if the input RNA was total RNA, an accurate calculation of fold-amplification requires knowledge of the Poly(A) content of the original total RNA sample. If the Poly(A) content of the sample is not known, a commonly used assumption is that Poly(A) RNA constitutes 2% of the RNA in a total RNA sample.

Fold-amplification = (amount of Biotin-aRNA produced) / ([amount of total RNA input] x [Percentage of Poly(A) RNA in the total RNA sample]).

Example: Amount of input total RNA = 200 ng, Percentage of Poly(A) RNA in the sample (assumed) = 2% (0.02), Amount of Biotin-aRNA produced = 30,000 ng (30 µg).

Fold-amplification = 30,000 ng / (200 ng x 0.02) = 7,500.

## F. Assessing the Size of the Biotin-aRNA Produced

Using the HeLa Control RNA provided in the kit, a TargetAmp 1-Round Biotin-aRNA Amplification Kit 105 reaction typically produces Biotin-aRNA with a size distribution between 200-4000 bases with an average size of about 800 bases.

Sizing the Biotin-aRNA by denaturing gel electrophoresis: The advantages of denaturing agarose gel electrophoresis are its relatively low cost and ready availability of the reagents required. When assessing the size distribution of the Biotin-aRNA, load at least 200 ng (SYBR® Gold detection) or at least 1 µg (ethidium bromide detection) of Biotin-aRNA into the well of a 1% formaldehyde-agarose gel. Load RNA size markers that cover the size range of approximately 200-10,000 bases.

*Sizing the Biotin-aRNA using the Agilent 2100 Bioanalyzer:* Use the appropriate molecular weight standards and follow the manufacturer's instructions.

## Abbreviated RNA Amplification Procedure for users experienced in using this kit.

### First-strand cDNA Synthesis (per reaction)

1. Anneal the TargetAmp T7-Oligo(dT) Primer A to the RNA sample.

x µl RNase-Free Water

x µl Total RNA sample

1 µl TargetAmp T7-Oligo(dT) Primer A

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3 µl Total reaction volume

2. Incubate at 65°C for 5 minutes, then chill on ice for 1 minute, then quick spin the sample.
3. Add 2 µl of the following 1<sup>st</sup>-Strand cDNA Synthesis Master Mix.

1.5 µl	TargetAmp Reverse Transcription PreMix
0.25 µl	DTT
0.25 µl	SuperScript III Reverse Transcriptase (200 U/µl)
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2 µl	Total reaction volume
4. Incubate at 50°C for 30 minutes.

**Second-strand cDNA Synthesis** (per reaction)

5. Add 5 µl of the following 2<sup>nd</sup>-Strand cDNA Synthesis Master Mix.

4.5 µl	TargetAmp DNA Polymerase PreMix
0.5 µl	TargetAmp DNA Polymerase
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5 µl	Total reaction volume
6. Incubate at 65°C for 10 minutes, quick spin the sample, then incubate at 80°C for 3 minutes, quick spin the sample then chill on ice.  
*In Vitro* Transcription of Biotin-aRNA (per reaction)
7. Add 20 µl of the following *In Vitro* Transcription Master Mix.

2 µl	T7 Transcription Buffer
3 µl	UTP / Biotin-UTP
10 µl	NTP PreMix
3 µl	DTT
2 µl	T7 RNA Polymerase
<hr/>	
20 µl	Total reaction volume
8. Incubate at 42°C for 4 hours.
9. Add 2 µl of RNase-Free DNase I, incubate at 37°C for 15 minutes.

**Biotin-aRNA Purification**

See Part D.

## 6. Appendix

### The TargetAmp 1-Round Biotin-aRNA Amplification Kit 105 Control Reaction

The TargetAmp kit provides 400 ng of total human HeLa RNA at a concentration of 40 ng/μl.

Required for the TargetAmp Control Reaction

Component Name	Tube Label	Cap Color
HeLa Total RNA Control (40 ng/μl)	HeLa Total RNA	Clear
RNase-Free Water	RNase-Free Water	
TargetAmp T7-Oligo(dT) Primer A	T7-Oligo(dT) Primer A	Red

1. Thaw the HeLa Total RNA Control on ice.
2. Anneal the TargetAmp T7-Oligo(dT) Primer A to the HeLa Total RNA Control.  
The standard control reaction utilizes 80 ng of the HeLa Total RNA Control.  
If using less Control RNA, make up the volume of the annealing reaction with RNase-Free Water.
 

2 μl	HeLa Total RNA Control (80 ng)
1 μl	TargetAmp T7-Oligo(dT) Primer A
3 μl	Total reaction volume
3. Incubate the reaction at 65°C for 5 minutes in a water bath or thermocycler. While the reaction incubates, quick-freeze the HeLa Total RNA Control (40 ng/μl; for example in a dry ice/ethanol bath) and return it to -70°C to -80°C storage.
4. Cool the annealing reaction on ice for at least 1 minute. Centrifuge the tube for 5-10 seconds to bring the sample to the bottom of the tube.
5. Continue the Control Reaction as described beginning in Part A, Step 4.

## 7. Additional TargetAmp aRNA Amplification Kits & Selection Guide

### TargetAmp™ 1-Round Aminoallyl-aRNA Amplification Kit 101

TAA1R4910 10 Reactions

TAA1R4924 24 Reactions

This kit will amplify Poly(A) RNA by >5,000 fold from as little as 25 ng of total cellular RNA. The kit produces aminoallyl-aRNA and is optimized for use of SuperScript III Reverse Transcriptase (provided by the user).

### TargetAmp™ 1-Round aRNA Amplification Kit 103

TAU1R5110 10 Reactions

TAU1R5124 24 Reactions

This kit will amplify Poly(A) RNA by >5,000 fold producing microgram amounts of unlabeled aRNA from as little as 25 ng of total cellular RNA. The kit is optimized for use with SuperScript III Reverse Transcriptase (provided by the user).

**TargetAmp™ 1-Round Biotin-aRNA Amplification Kit 104**

TAB1R6910 10 Reactions

TAB1R6924 24 Reactions

This kit will amplify Poly(A) RNA by >5,000 fold from as little as 25 ng of total cellular RNA. The kit produces biotin-aRNA and is optimized for use of SuperScript III Reverse Transcriptase (provided by the user).

**TargetAmp™ Nano Labeling Kit for Illumina® Expression BeadChip®**

TAN07924 24 Reactions

This kit will produce microgram amounts of Biotin-aRNA from as little as 25 ng of total cellular RNA for hybridization to Illumina® Expression BeadChips. The kit is optimized for use with SuperScript III Reverse Transcriptase (provided by the user).

**TargetAmp™ 2-Round Aminoallyl-aRNA Amplification Kit 1.0**

TAA2R4910 10 Reactions

TAA2R4924 24 Reactions

This kit will amplify Poly(A) RNA by >5,000,000 fold producing microgram amounts of aminoallyl-aRNA from as little as 10 pg of total cellular RNA. The kit is optimized for use with SuperScript III and SuperScript II Reverse Transcriptases (provided by the user).

**TargetAmp™ 2-Round aRNA Amplification Kit 2.0**

TAU2R5110 10 Reactions

TAU2R51224 24 Reactions

This kit will amplify Poly(A) RNA by >5,000,000 fold producing microgram amounts of unlabeled aRNA from as little as 10 pg of total cellular RNA. The kit is optimized for use with SuperScript III and SuperScript II Reverse Transcriptases (provided by the user).

**TargetAmp™ 2-Round Biotin-aRNA Amplification Kit 3.0**

TAB2R71010 10 Reactions

TAB2R71024 24 Reactions

This kit will amplify Poly(A) RNA by >5,000,000 fold producing microgram amounts of biotin-aRNA from as little as 50 pg of total cellular RNA. The kit is optimized for use with SuperScript III and SuperScript II Reverse Transcriptases (provided by the user).

	TargetAmp™ 1-Round Aminoallyl-aRNA Amplification Kit 101	TargetAmp™ 1-Round aRNA Amplification Kit 103	TargetAmp™ 1-Round Biotin-aRNA Amplification Kit 105	TargetAmp™ Nano Labeling Kit for Illumina® Expression BeadChip®
Starting Total RNA	25-500 ng	25-500 ng	25-500 ng	25-500 ng
Reverse Transcriptase(s) Used	SuperScript® III (provided by the user)	SuperScript® III (provided by the user)	SuperScript® III (provided by the user)	SuperScript® III (provided by the user)
Fold Amplification	> 5,000	> 5,000	> 5,000	> 5,000
Time Required	1 Day	1 Day	1 Day	1 Day
End Product	Aminoallyl-aRNA	Unlabeled-aRNA	Biotin-aRNA	Biotin-aRNA

	TargetAmp™ 2-Round Aminoallyl-aRNA Amplification Kit 1.0	TargetAmp™ 2-Round aRNA Amplification Kit 2.0	TargetAmp™ 2-Round Biotin-aRNA Amplification Kit 3.0
Starting Total RNA	10-500 ng	10-500 pg	50-500 pg
Reverse Transcriptase(s) Used	SuperScript® III & SuperScript II (provided by the user)	SuperScript® III & SuperScript II (provided by the user)	SuperScript® III & SuperScript II (provided by the user)
Fold Amplification	>5,000,000	>5,000,000	>5,000,000
Time Required	2 Days	2 Days	2 Days
End Product	Aminoallyl-aRNA	Unlabeled-aRNA	Biotin-aRNA

## 8. Reference

1. Van Gelder, R. N. *et al.*, (1990) *Proc. Natl. Acad. Sci. USA* **87** (5), 1663.

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