

# CopyControl™ pCC1BAC™ (*Bam*H I, *Hind* III, *Eco*R I Cloning-Ready) Vector

Cat. Nos. CBAC311B, CBAC311H, and CBAC311E

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## 1. Introduction

The CopyControl™ pCC1BAC™ Vector\* is based on an innovative technology originally developed in the laboratory of Dr. Waclaw Szybalski<sup>1</sup> and optimized at Epicentre.<sup>2</sup> The vector has two origins of replication – a single-copy *E. coli* F-factor replicon and a high-copy origin of replication called "*oriV*". Initially, replication of CopyControl clones can be controlled by the F-factor replicon so the vector is present at one copy per cell. Maintaining clones at single copy ensures insert stability and allows cloning of toxic gene products (Fig. 1).

Initiation of replication from *oriV* requires the *trfA* gene product. CopyControl Vectors use a specifically engineered *E. coli* host strain, TransforMax™ EPI300™ (available separately), which contains a mutant *trfA* gene under tight control of an inducible promoter. Addition of the CopyControl Induction Solution to the growth medium induces expression of *trfA* and subsequent amplification of the clone to high-copy number. Induction of CopyControl BAC clones from single-copy up to 10-20 copies per cell greatly improves the yield and purity of BAC DNA for sequencing, fingerprinting and other applications.

The CopyControl pCC1BAC Vector is derived from pBeloBAC113 and Epicentre's pIndigoBAC-5. The vector has been linearized at a unique restriction enzyme recognition site (*Bam*H I, *Hind* III or *Eco*R I), dephosphorylated, and highly purified to ensure very low background. Features of the vector include:

- Chloramphenicol-resistance as an antibiotic selectable marker.
- *E. coli* F factor-based partitioning and single-copy number regulation system.
- *oriV* high-copy origin of replication.
- Primer binding sites for BAC-end sequencing
- *Not* I sites surrounding the *Bam*H I, *Hind* III and *Eco*R I cloning sites.
- Bacteriophage P1 *loxP* site for Cre-recombinase cleavage.

## 2. Product Specifications

**Storage:** Store at –20°C.

**Size:** 375 ng @ 25 ng/μl (15 μl) (in TE Buffer (10 mM Tris-HCl [pH 7.5], 1 mM EDTA)

**Quality Control:** Cloning-ready preparations of the CopyControl pCC1BAC Vector yield >10<sup>7</sup> cfu/μg of Control Insert DNA when transformed into TransforMax EPI300 Electrocompetent *E. coli*. Greater than 95% of the colonies are recombinant clones.

**Protocols:** See References 4-7 for protocols on BAC cloning and working with BAC clones. Product literature for the CopyControl BAC Cloning Kits also provides thorough procedures for constructing a BAC library.

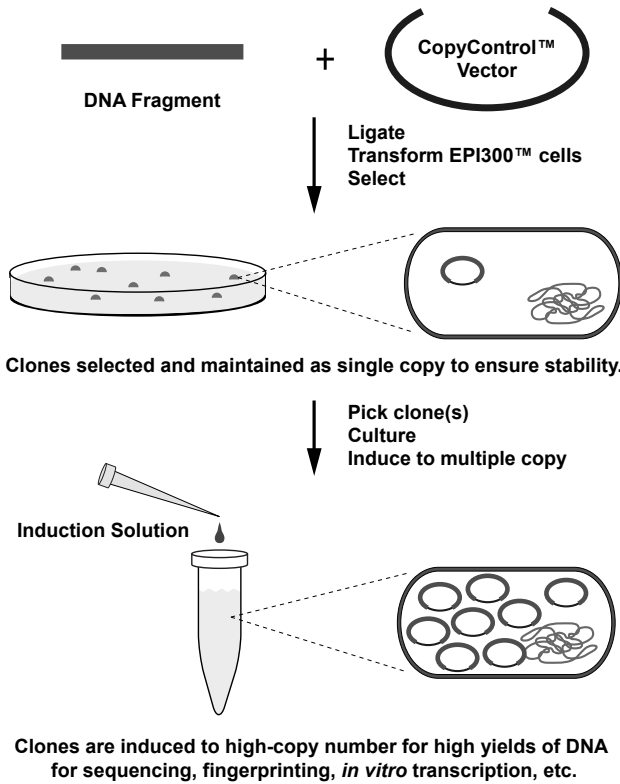
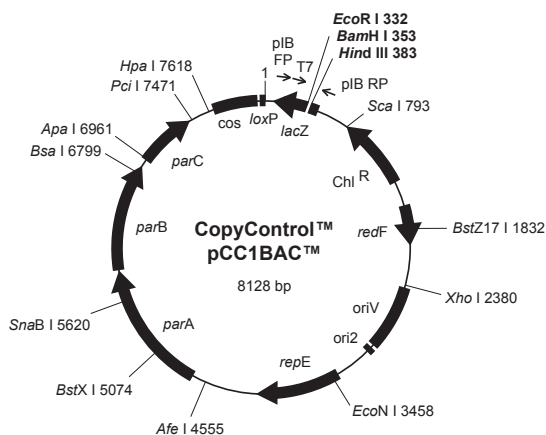


Figure 1. Overview of the CopyControl™ System.

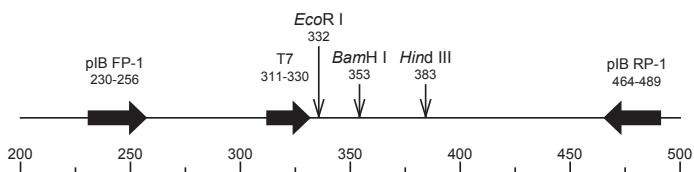
### 3. Related Products

The following products are also available:

- CopyControl™ BAC Cloning Kits
- TransforMax™ EPI300™ Electrocompetent and Chemically Competent *E. coli*
- TransforMax™ EPI300™-T1<sup>R</sup> Electrocompetent and Chemically Competent *E. coli*
- BACMAX™ DNA Purification Kit
- Fast-Link™ DNA Ligation Kits
- Colony Fast-Screen™ Kit (Size Screen)
- EZ-Tn5™ <*ori*V/KAN-2> Insertion Kit
- GELase™ Gel-Digesting Preparation
- Plasmid-Safe™ ATP-Dependent DNase



Note: Not all restriction enzymes that cut only once are indicated above. See page 6 for complete restriction information. Primers are not drawn to scale.



FP = pCC1™/pEpiFOS™ Forward Sequencing Primer 5' GGATGTGCTGCAAGCGGATTAAGTTGG 3'  
 RP = pCC1™/pEpiFOS™ Reverse Sequencing Primer 5' CTCGTATGTTGTGTGGAATTGTGAGC 3'  
 T7 = T7 Promoter Primer 5' TAATACGACTCACTATAGGG 3'

Figure 2. CopyControl™ pCC1BAC™ Vector.

#### 4. How the CopyControl Cloning System Works

1. Ligate the DNA interest into the linearized and dephosphorylated CopyControl pCC1 Cloning-Ready Vector.
2. Transform TransformMax EPI300 Electrocompetent *E. coli* and select clones on LB-chloramphenicol plates. Under these conditions, the *trfA* gene is repressed and the clones are maintained as single copy.
3. Pick individual CopyControl clones from the plate and grow in culture.
4. Add the CopyControl Induction Solution to induce expression of the *trfA* gene product and subsequent amplification of the clones to high copy number.
5. Purify plasmid DNA for sequencing, fingerprinting, subcloning or other applications.

**Important:** An *E. coli* host carrying an inducible *trfA* gene (such as TransformMax EPI300 *E. coli* or phage T1-resistant TransformMax EPI300-T1<sup>R</sup> *E. coli*) is required for amplification of the CopyControl BAC clones to high-copy number. A regulated *trfA* gene is not present in most lab strains of *E. coli*. We can not guarantee clone amplification results using any *E. coli* strain other than TransformMax EPI300 *E. coli*, which are available separately.

## 5. pCC1BAC Sequencing Primers and Vector Data

### pCC1 / pEpiFOS-5 Sequencing Primers

pCC1™ / pEpiFOS™ Forward Sequencing Primer ..... Cat. No. F5FP010

5' GGATGTGCTGCAAGGCGATTAAGTTGG 3' ..... 1 nmol supplied in TE Buffer at 50 μM

pCC1™ / pEpiFOS™ Reverse Sequencing Primer ..... Cat. No. F5RP011

5' CTCGTATGTTGTGTGGAATTGTGAGC 3' ..... 1 nmol supplied in TE Buffer at 50 μM

### pCC1 / pEpiFOS Forward Sequencing Primer

#### Temperatures of Dissociation & Melting:

$T_d$ : 79°C (nearest neighbor method)

$T_m$ : 78°C (% G+C method)

$T_m$ : 82°C ([2 (A+T) + 4 (G+C)] method)

$T_m$ : 68°C ((81.5 + 16.6 (log [Na<sup>+</sup>])) +  
([41 (#G+C) - 500] / length) method)  
where [Na<sup>+</sup>] = 0.1 M

### pCC1 / pEpiFOS Reverse Sequencing Primer

#### Temperatures of Dissociation & Melting:

$T_d$ : 71°C (nearest neighbor method)

$T_m$ : 75°C (% G+C method)

$T_m$ : 76°C ([2 (A+T) + 4 (G+C)] method)

$T_m$ : 65°C ((81.5 + 16.6 (log [Na<sup>+</sup>])) +  
([41 (#G+C) - 500] / length) method)  
where [Na<sup>+</sup>] = 0.1 M

**Note:** The sequence of the pCC1/pEpiFOS Forward and Reverse Primers do not function well as IRD800-labeled sequencing primers. We recommend using the T7 and pCC1/pEpiFOS RP-2 Primers instead of the pCC1/pEpiFOS Forward and Reverse Primers respectively, for this purpose.

### pCC1™ / pEpiFOS™ RP-2 Reverse Sequencing Primer

5' TACGCCAAGCTATTTAGGTGAGA 3'

### Orientation for BAC End-Sequencing

The following is the nucleotide sequence of pCC1BAC (bases 230-489) from the pCC1/pEpiFOS Forward Sequencing Primer (230-256) to the pCC1/pEpiFOS Reverse Sequencing Primer (489-464) encompassing the T7 RNA polymerase promoter (311-330) the *Eco*R I site (332-337), the *Bam*H I site (353-358) and the *Hind* III site (383-388).

```
230 GGATGTGCTG CAAGGCGATT AAGTTGGGTA ACGCCAGGGT TTTCCCAGTC  
280 ACGACGTTGT AAAACGACGG CCAGTGAATT GTAATACGAC TCACTATAGG  
330 GCGAATTCGA GCTCGGTACC CGGGGATCCT CTAGAGTCGA CCTGCAGGCA  
380 TGCAAGCTTG AGTATCTTAT AGTCTCACCT AAATAGCTTG GCGTAATCAT  
430 GGTCATAGCT GTTTCTGTG TGAAATTGTT ATCCGCTCAC AATTCCACAC  
480 AACATACGAG
```

An electronic copy of the pCC1BAC sequence is available for downloading at our Web site at <http://www.epicentre.com.sequences> or can be requested via e-mail ([techhelp@epicentre.com](mailto:techhelp@epicentre.com)) or by calling Technical Service.

**Restriction Enzymes that cut the pCC1BAC Vector one to three times:**

| <b>Enzyme</b> | <b>Sites</b> | <b>Location</b>  | <b>Enzyme</b> | <b>Sites</b> | <b>Location</b>  |
|---------------|--------------|------------------|---------------|--------------|------------------|
| Acc65 I       | 2            | 344, 5196        | Fsp I         | 3            | 167, 3741, 7567  |
| Acl I         | 2            | 1121, 5588       | Hind III      | 1            | 383              |
| Afe I         | 1            | 4555             | Hpa I         | 1            | 7618             |
| Afl II        | 2            | 6597, 6837       | Kpn I         | 2            | 348, 5200        |
| Afl III       | 3            | 4962, 5136, 7471 | Mfe I         | 1            | 4976             |
| Age I         | 3            | 3816, 5046, 5939 | Msc I         | 3            | 943, 2623, 5407  |
| Ahd I         | 1            | 7475             | Nar I         | 1            | 146              |
| Ale I         | 1            | 6532             | Nco I         | 2            | 905, 7176        |
| Apa I         | 1            | 6961             | Nde I         | 2            | 94, 4994         |
| ApaB I        | 3            | 96, 1934, 7635   | Not I         | 2            | 2, 631           |
| ApaL I        | 1            | 87               | Nru I         | 2            | 1632, 7663       |
| BamH I        | 1            | 353              | Nsp I         | 3            | 381, 1819, 7475  |
| Bbs I         | 3            | 5039, 5228, 6105 | PaeR7 I       | 1            | 2380             |
| BciV I        | 1            | 2486             | Pci I         | 1            | 7471             |
| Bcl I         | 1            | 5787             | PflF I        | 1            | 5260             |
| Bgl I         | 3            | 639, 3160, 7609  | PpuM I        | 2            | 1716, 7847       |
| Bgl II        | 2            | 3135, 5202       | Psi I         | 2            | 2915, 3111       |
| Blp I         | 1            | 4468             | PspOM I       | 1            | 6957             |
| BmgB I        | 3            | 2613, 5026, 7786 | Pst I         | 3            | 375, 4014, 5555  |
| Bmr I         | 3            | 268, 7007, 7136  | Pvu I         | 2            | 188, 5862        |
| Bpu10 I       | 3            | 1434, 3916, 5111 | Sac II        | 1            | 2472             |
| Bsa I         | 1            | 6799             | Sal I         | 3            | 365, 645, 7651   |
| BsaB I        | 2            | 7743, 7827       | Sap I         | 2            | 4592, 5802       |
| BsaH I        | 1            | 146              | Sbf I         | 2            | 375, 4014        |
| BseY I        | 3            | 2401, 5879, 6636 | Sca I         | 1            | 793              |
| Bsm I         | 2            | 812, 1219        | SexA I        | 1            | 7589             |
| BsmB I        | 3            | 982, 1535, 3931  | Sfi I         | 1            | 639              |
| BspE I        | 2            | 1210, 5756       | Sfo I         | 1            | 147              |
| BspLU11 I     | 1            | 7471             | SgrA I        | 3            | 2481, 5046, 6203 |
| BsrB I        | 3            | 464, 1648, 2270  | Sim I         | 2            | 5160, 7847       |
| BsrG I        | 1            | 3769             | Sma I         | 3            | 350, 639, 3482   |
| BssH II       | 2            | 5453, 5997       | SnaB I        | 1            | 5620             |
| BssS I        | 3            | 5146, 6796, 7359 | Spe I         | 1            | 6711             |
| BstAP I       | 3            | 95, 1933, 7634   | Sph I         | 1            | 381              |
| BstE II       | 1            | 7593             | Srf I         | 1            | 639              |
| BstX I        | 1            | 5074             | Sse8647 I     | 1            | 1716             |
| BstZ17 I      | 1            | 1832             | Stu I         | 1            | 3163             |
| Bts I         | 2            | 558, 5548        | Tat I         | 3            | 77, 791, 3769    |
| Dra III       | 2            | 1933, 7812       | Tli I         | 1            | 2380             |
| Eco47 III     | 1            | 4555             | Tth111 I      | 1            | 5260             |
| EcoN I        | 1            | 3458             | Xba I         | 2            | 359, 3181        |
| EcoO109 I     | 2            | 1716, 7847       | Xcm I         | 1            | 2676             |
| EcoR I        | 1            | 332              | Xho I         | 1            | 2380             |
| EcoRV         | 2            | 4117, 4346       | Xma I         | 3            | 348, 637, 3480   |
| Fse I         | 1            | 2478             |               |              |                  |

**Restriction Enzymes that cut the pCC1BAC Vector four or more times:**

|           |           |            |          |          |
|-----------|-----------|------------|----------|----------|
| Acc I     | BsmA I    | Dsa I      | HpyCH4 V | PspG I   |
| Aci I     | Bsp1286 I | Eae I      | Mae II   | Pvu II   |
| Alu I     | BspH I    | Eag I      | Mae III  | Rsa I    |
| Alw I     | BspM I    | Ear I      | Mbo I    | Sac I    |
| AlwNI     | Bsr I     | Fau I      | Mbo II   | Sau3A I  |
| Apo I     | BsrD I    | Fnu4H I    | Mly I    | Sau96 I  |
| Ase I     | BsrF I    | Gdi II     | Mnl I    | ScrF I   |
| Ava I     | BssKI     | Hae I      | Mse I    | SfaNI    |
| Ava II    | BstDS I   | Hae II     | Msl I    | Sfc I    |
| Ban I     | BstF5 I   | Hae III    | Msp I    | Sml I    |
| Ban II    | BstNI     | Hha I      | MspA1 I  | Ssp I    |
| Bfa I     | BstU I    | Hinc II    | Mwo I    | Sty I    |
| BfuA I    | BstY I    | Hinf I     | Nae I    | Taq I    |
| Bme1580 I | Btg I     | HinP I     | Nci I    | Tfi I    |
| BsaA I    | Cac8 I    | Hpa II     | NgoM IV  | Tse I    |
| BsaJ I    | CviJ I    | Hph I      | Nla III  | Tsp45 I  |
| BsaW I    | Dde I     | Hpy188 I   | Nla IV   | Tsp4C I  |
| BsiE I    | Dpn I     | Hpy99 I    | PflM I   | Tsp509 I |
| BsiHKA I  | Dra I     | HpyCH4 III | Ple I    | TspR I   |
| Bsl I     | Drd I     | HpyCH4 IV  | PshA I   | Xmn I    |

**Restriction Enzymes that do not cut the pCC1BAC Vector:**

|        |        |         |       |        |
|--------|--------|---------|-------|--------|
| Aat II | BbvC I | BstB I  | Nhe I | Pml I  |
| Asc I  | BfrB I | Bsu36 I | Nsi I | Rsr II |
| AsiS I | BsiW I | Cla I   | Pac I | SanD I |
| Avr II | BspD I | Mlu I   | Pme I | Swa I  |



## 6. References

1. Wild, J. *et al.*, (2002) *Genomic Research* **12**, 1434.
2. *Epicentre Forum* (2002) **9** (1), 1.
3. Hurowitz, E.H. *et al.*, (2000) *DNA Research* **7** (2), 1.
4. Birren, B. *et al.*, (1999) Bacterial Artificial Chromosomes in *Genome Analysis: A Laboratory Manual*, CSH Press, New York, **v. 3**, 241.

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